Morphology of COVID-19–affected cells in peripheral blood film

Aminder Singh, Neena Sood, Vikram Narang, Abhishek Goyal

DESCRIPTION
A 55-year-old previously healthy woman was admitted with fever and cough. Chest X-ray and CT showed features of viral pneumonitis. Her nasopharyngeal swab was positive for severe acute respiratory distress syndrome coronavirus-2 (SARS-CoV-2) by reverse transcription (RT)-PCR. Her complete blood count (CBC) showed leukocytosis with neutrophilia, relative lymphocytopenia, and monocytopenia initially with subsequent improvement in the number of monocytes on the fifth day onwards. We report a detailed analysis of the peripheral blood film (PBF) of a patient with COVID-19 during the hospital course which has not been reported hitherto. Some peculiar findings were observed in the neutrophils which were never seen in any other infection. Neutrophils showed heavily clumped chromatin with toxic granules and cytoplasmic vacuoles. Nuclear detachment with elongated nucleoplasm and ring-shaped nuclei were seen with platelet surface attachment. C-shaped, fetus-like nuclei were noted with aberrant nuclear projections, which we named as COVID nuclei (figure 1). Most of the lymphocytes were seen as large granular lymphocytes (LGL) with round to indented nuclei, condensed chromatin, prominent nucleoli in a few, along with abundant pale blue cytoplasm with distinct variably sized azurophilic granules (figure 2). Cytoplasmic pod formation and apoptosis were also observed in a few lymphocytes. These might represent natural killer cells or cytotoxic T lymphocytes. Activated monocytes were seen which showed marked anisocytosis with prominent cytoplasmic vacuolisation and few granules. Nuclei were large, having fine chromatin with nuclear blebbing in a few. Nuclear overlapping by vacuoles was observed in some cells (figure 3). Platelets were adequate, with a few giant forms and focal platelet attachment on the surface of

Figure 1 Peripheral blood films showing various neutrophils with C-shaped, fetus-like COVID nuclei (black arrowheads) with aberrant nuclear projections (blue arrowhead). Toxic granulations and vacuolations (yellow arrowhead), ring nuclei (red arrowheads) and elongated nucleoplasm (green arrowheads) are highlighted. Giemsa ×200–400.

Figure 2 Peripheral blood films showing large granular lymphocytes. Round to indented nuclei, condensed chromatin, prominent nucleoli in a few, along with abundant pale blue cytoplasm with distinct variably sized azurophilic granules are present (long black arrow). Cytoplasmic pod formation (long green arrows) and apoptotic lymphocytes (long red arrow) are highlighted. Giemsa ×200–400.

Figure 3 Peripheral blood films showing activated monocytes with prominent cytoplasmic vacuolisation and a few granules (small red arrow). Nuclear blebbing (small green arrow) is also seen. Giemsa ×200–400.
all forms of leucocytes. Her first CBC sample showed normal total leucocyte count (7.4×10⁹/L) with neutrophilia and relative lymphocytopenia and monocytopaenia. On the fifth day onwards, her CBC started showing monocytopaosis, which persisted until day 16. Relative lymphocytopenia persisted throughout her stay, along with the presence of LGL on PBF. LGL started appearing on PBF on day 7 onwards. These findings clearly indicated that in COVID-19, initial neutrophilia, lymphocytopenia and monocytopaenia are subsequently accompanied by monocytosis. During the hospital stay, the patient was managed with oral hydroxychloroquine, azithromycin and antipyretics, along with intravenous fluids, bronchodilators and supportive care. The patient was discharged on 8 April 2020 after two negative samples for COVID-19.

COVID-19 is a highly contagious disease caused by SARS-CoV-2. It originated from the Chinese city of Wuhan and has reached the pandemic status in just a few months. It has now affected around 300 countries worldwide, causing significant morbidity and mortality. To date more than 254,045 deaths have been reported.¹

There are various challenges to the exact diagnosis of these patients. Clinically patients may be asymptomatic carriers. Sensitivity and specificity of currently available methods are very variable.²

We have examined the PBF findings in a patient with COVID-19 which can help in the diagnosis and to some extent the prognosis of patients. The inflammatory response and viral effects on leucocytes might be responsible for these changes, which can be readily identified on PBF and can be easily and serially monitored. Monocytosis may be associated with favourable outcome, as seen in our patient.³

Understanding of the haematological manifestations of SARS-CoV-2 is still in the evolving stage. To reach a definite conclusion regarding the specificity and reliability of these viral cytopathic effects in peripheral smear, more patients positive for COVID-19 need to be evaluated in larger studies. Our goal is to describe the peculiar morphological findings of affected leucocytes, which would help physicians suspect a diagnosis in the absence of a negative RT-PCR or antibody results. If confirmed in larger studies, these morphological features along with blood count would be helpful in the screening, diagnosis and management of these patients at all levels of healthcare.

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